

Slice SILAC : coupling isotope labeling with electrophoretic migration to study protein processing events.

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INTRODUCTION

Metabolic isotope labeling of proteins (e.g. SILAC) is commonly used in conjunction with LC-MS/MS for quantitative proteomics. To decrease complexity, samples are often fractionated at the peptide level after total digestion. Alternatively, 1D SDS-PAGE can be used to separate proteins based on molecular weight, followed by in-gel digestion of gel slices (geLC-MS workflow).

While showing some drawbacks for global quantification, the geLC-MS workflow allows to retain information on protein mass and thus has the potential to reveal post-translational processing events normally invisible with a total digestion / peptide fractionation approach.

We analysed 1D gel-separated SILAC mixtures of lysates from **heat-shocked** (44C) vs. control Jurkat cells. Data were analysed with MaxQuant (1) first by global quantification then slice-by-slice and the results were compared.

CONCLUSIONS

Compilation of spectral counts as a function of gel slice provided a low resolution *virtual western blot* profile for most (>1400) identified proteins. The addition of SILAC ratio information per slice allows to detect **changes in protein mobility** induced by the treatment. We detected significant heat shock-induced mobility shifts for 35 proteins. The pattern detected was consistent with a proteolytic cleavage for 32 proteins, of which 22 were previously known caspase substrates (2), thus revealing the onset of an apoptotic cascade.

Therefore a **slice SILAC** analysis can offer a completely different image (**more based on post-translational modifications**) of the cellular reaction to a stimulus than the one obtained with a global protein quantitation analysis.

